

Nematostatic N₂-fixing *Crotalaria* for greenhouse vegetable production (GVP) in Mediterranean conditions

Philippe de Lajudie

International Centre in Agronomic Research and Development (CIRAD), France

AABNF Rabat November 2014



Legume-supported cropping systems for Europe (Legume Futures)
is a collaborative research project funded from the European Union's Seventh Programme
for research, technological development and demonstration under grant number 245216

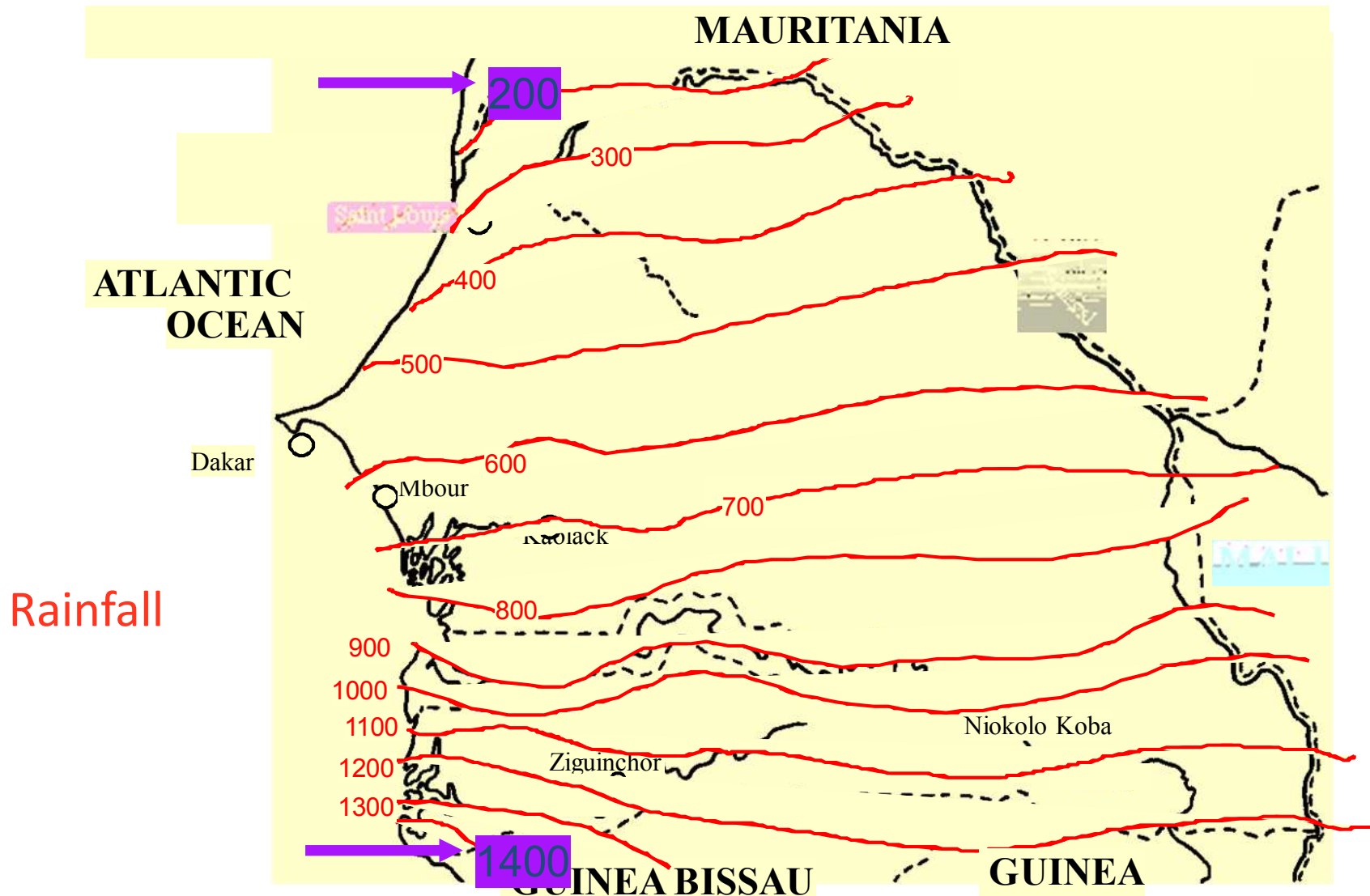
www.legumefutures.de

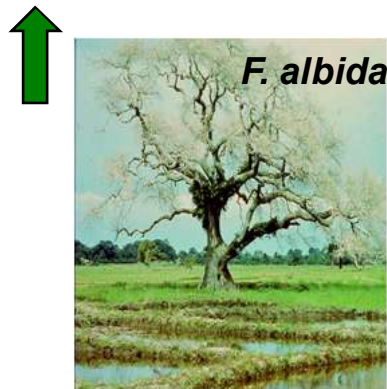
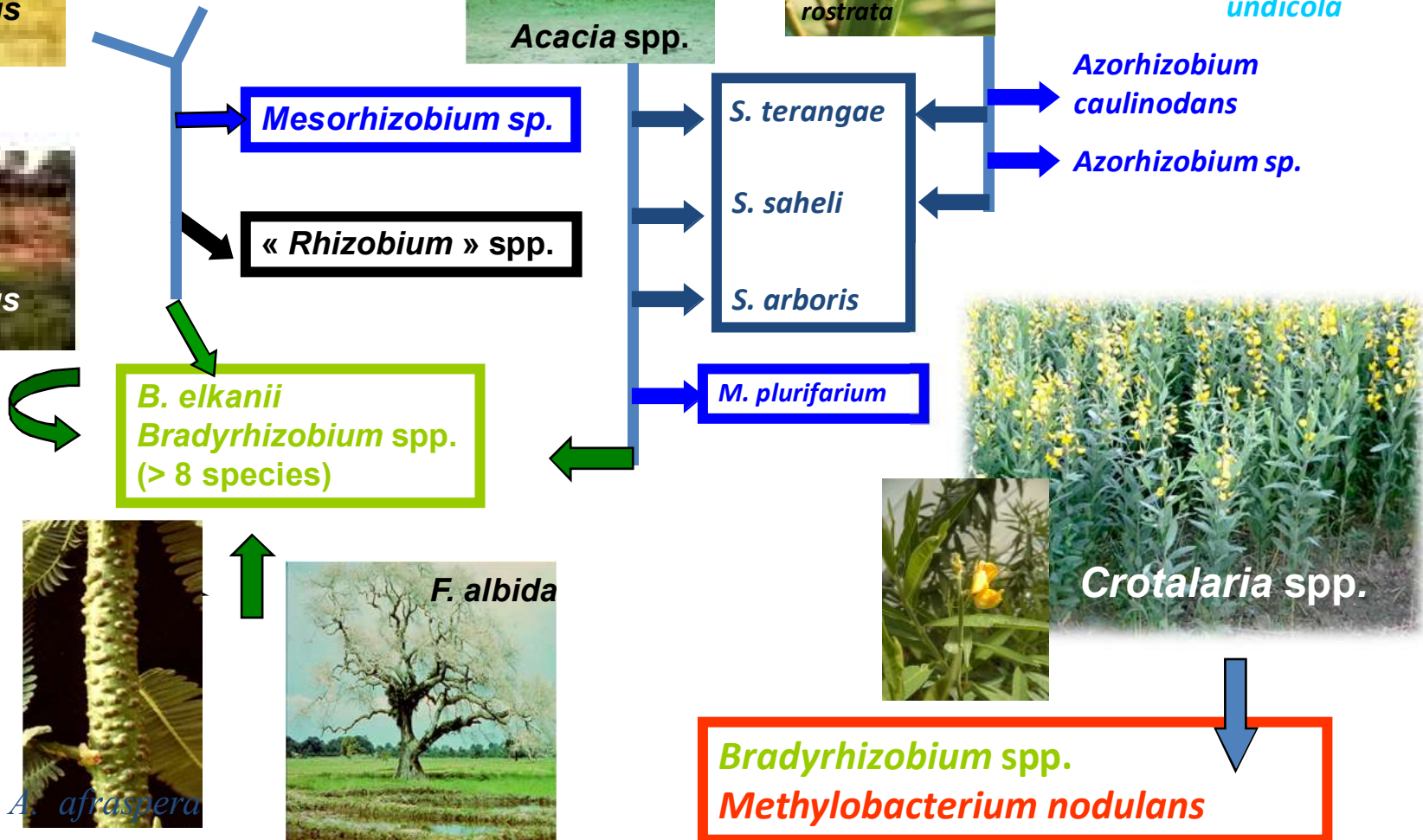


Tunnel
in
South
France

Canarian shelter in Spain

Investigating and promoting local legume symbioses for development in Senegal



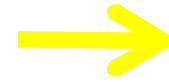


de Lajudie et al., 1994;1998ab;

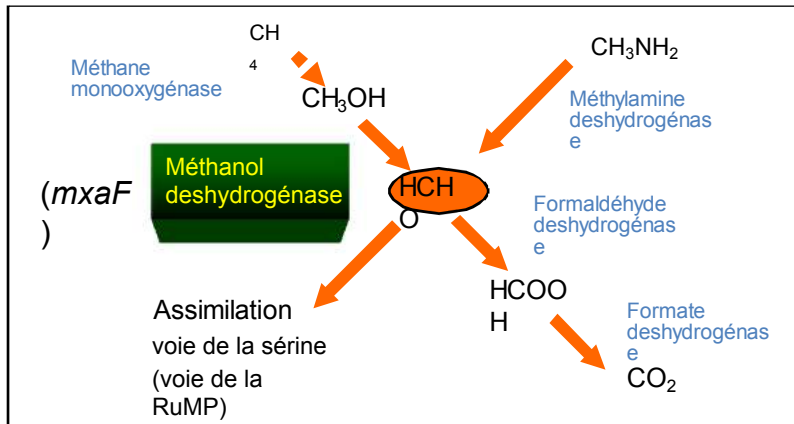
Diouf et al., 1999 ; 2007; Doignon-Bourcier et al., 1999;2000;

Sylla et al., 2002; Sy et al., 2001; Jourand et al., 2004

Investigating local legume symbioses



Unexpected properties in LNBs



New models

Potential of Legume symbioses

- Plant nutrition (BNF, P)
- Metabolite production
- Land reclamation
- Crop protection (parasites)
- ...

Méthylotrophy

Biodegradation

Carotenoids

Photosynthesis

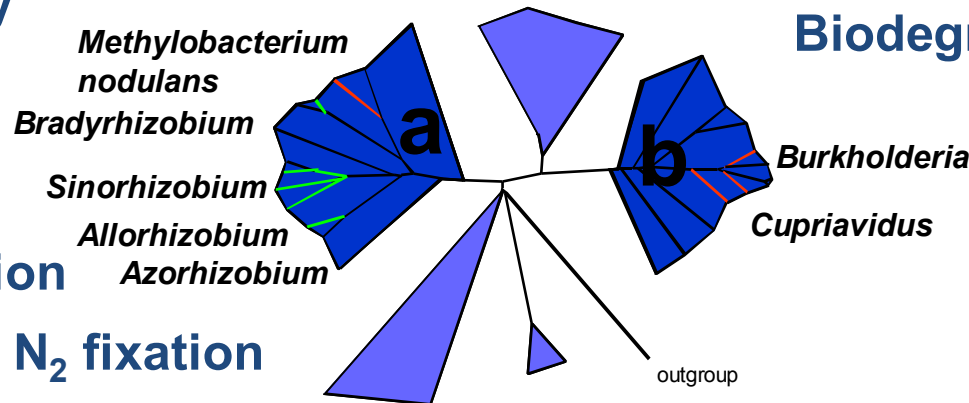
Stem nodulation

Free-living N_2 fixation

nod gene independant nodulation

(endo)colonization of non-legumes: rice, maize

P solubilisation (FABATROPIMED)



Dreyfus et al., 1981
 Lorquin et al., 1997
 Molouba et al., 1999
 Chaintreuil et al., 2000
 Giraud et al., 2000, 2002
 Sy et al., 2001
 Moulin et al., 2001
 Chen et al., 2001
 Jourand et al., 2004, 2005
 N'Zoué, 2008
 Merabet et al., 2010

**No efficient treatment against nematodes.
Tourteau of castor-oil plant reputed active
Common practice: 3 month sanitary period (fallow)
Solarization / xolone treatment**



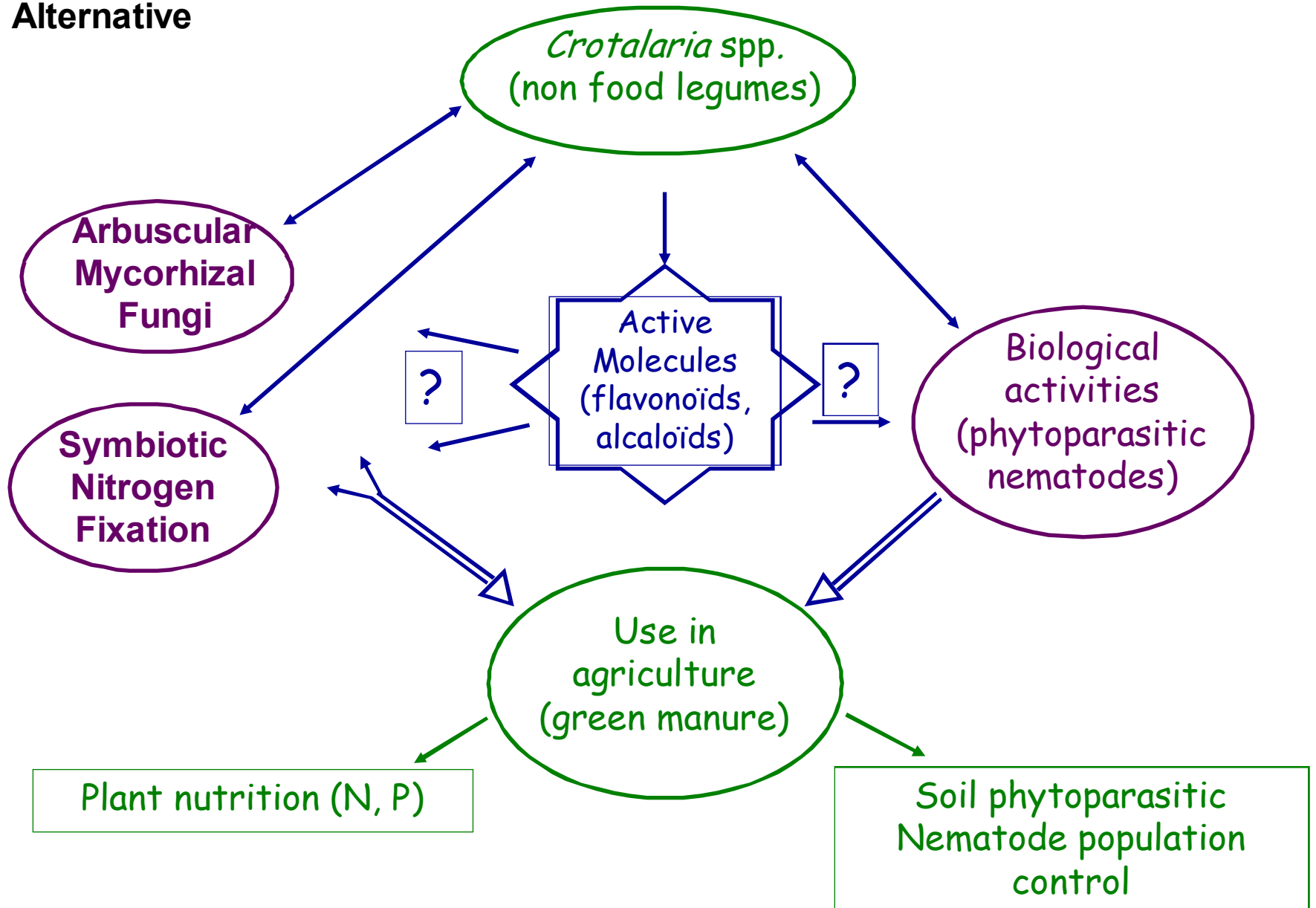
Chemicals against nematodes are:

- toxic to the users
- soon banned in Europe

**Alternative
Non-food legume *Crotalaria***



Alternative

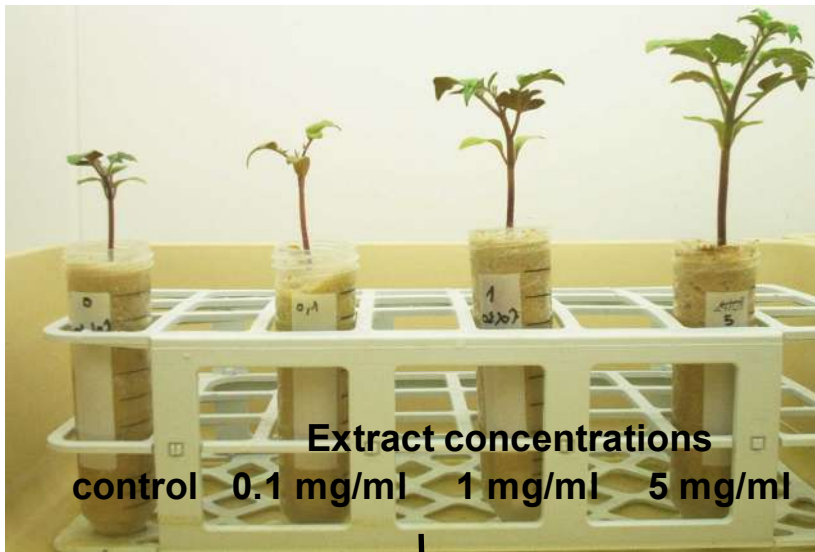


Crotalaria Nematostatic Activity

Early data

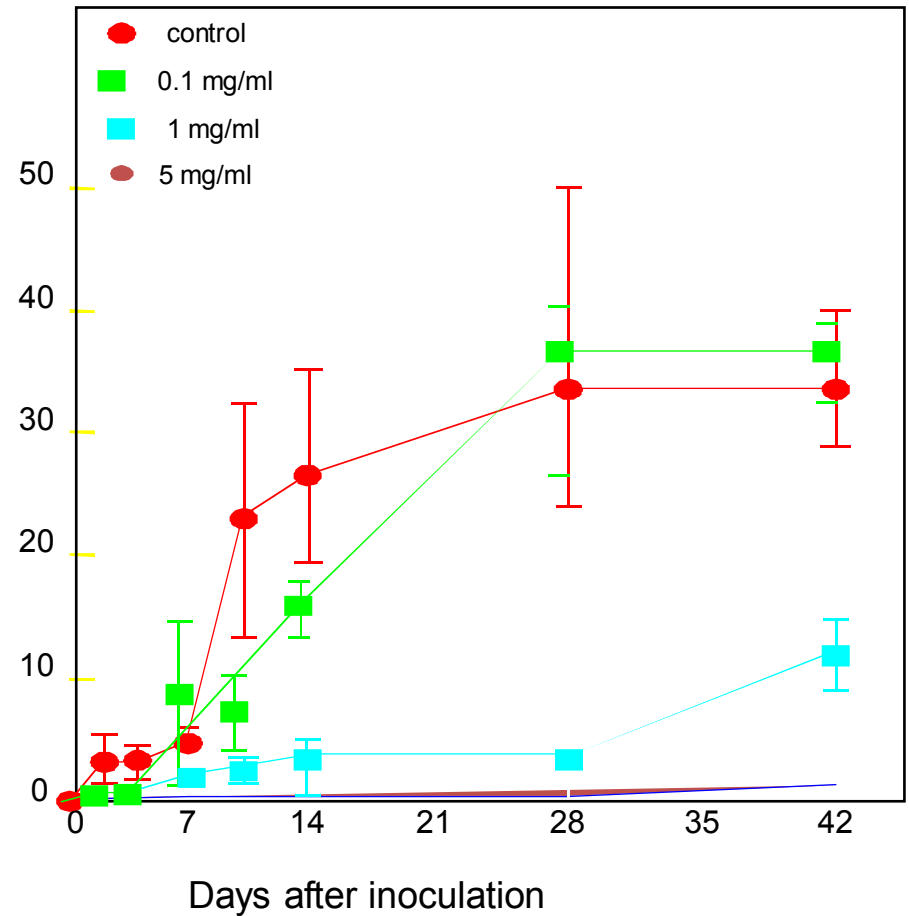
Tomato Plants
+ *Crotalaria grantiana*
foliar extracts

Inoculated with 100 *M. incognita* juveniles



% root infestation
fuschine coloration

% juveniles infested roots



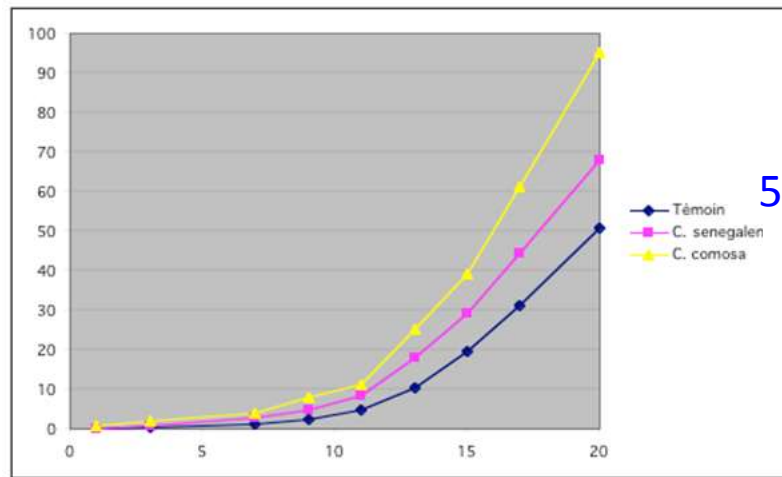
Phytoprotection effect : delay and infestation inhibition

	<i>Meloïdogyne incognita</i>		<i>Meloïdogyne javanica</i>	
	Penetration rate	Dev to adult	Penetration rate	Dev to Adult
<i>Crotalaria</i> (15 species)	0,2 - 7% 0%	0,6 - 5% 0%		
Tomato	97%	74%	77%	13%

Jourand, 2004

Nematostatic effect of *Crotalaria*

Early data in LCM, Dakar, Senegal



Tomato yield (kg / 20 plants)



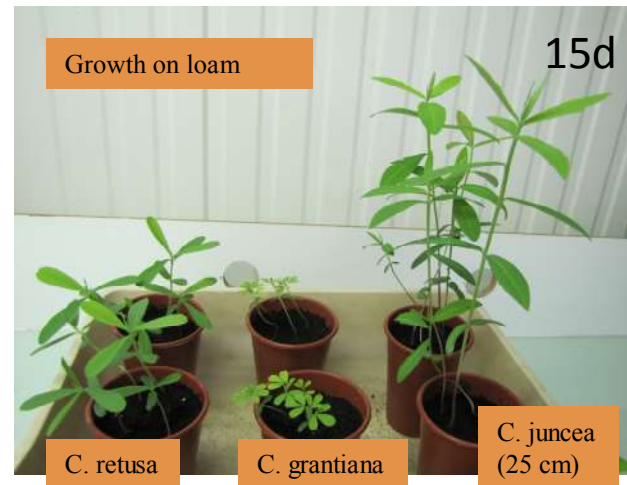
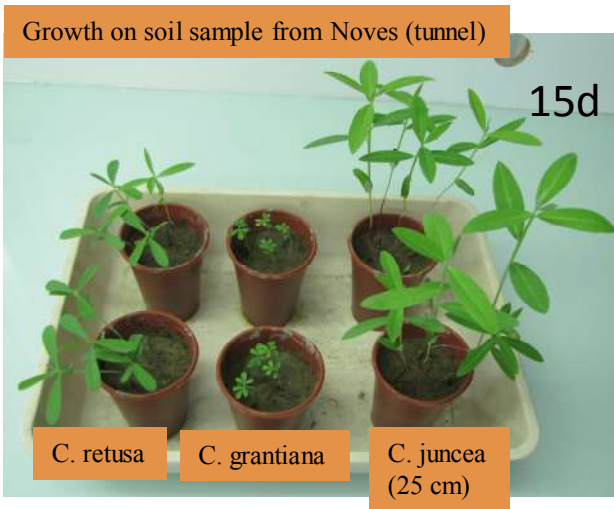
Control

Temps (j)

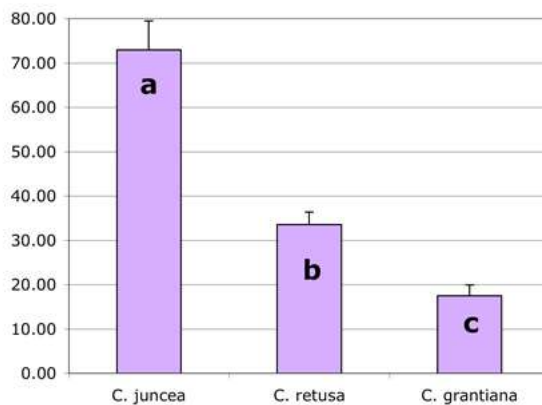
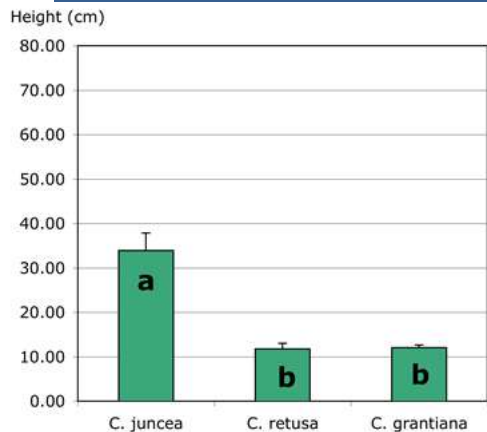
LSTM Objectives

To test *Crotalaria* spp. for **plant protection** against nematodes as intercrop/rotational **green manure in industrial greenhouse vegetable production** on the commercial scale for a cost-effective, environment-friendly and **sustainable agriculture in 3 Mediterranean countries** with the private company Delbon

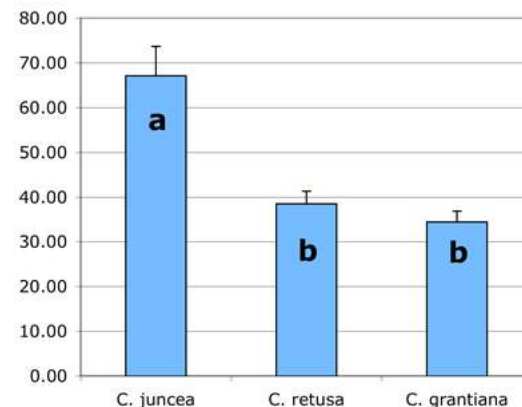
Comparative growth of 3 *Crotalaria* species on 3 substrates



Crotalaria juncea produces significant more biomass



Height of 3 *Crotalaria* species after 3 months of growing in pots filled with loam in greenhouse



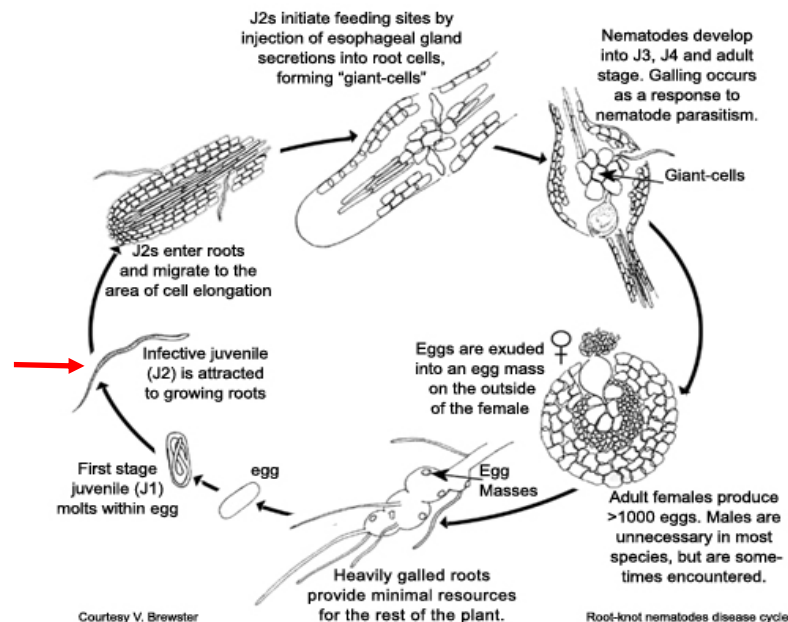
Tropical **Shrub** Legume (1-4m ; *Fabaceae*, *Papilionoidae*, *Crotalariae*)

Not edible. Cultivated in India for **fibers and rope** production

Nitrogen Fixing symbiosis

Highly **mycotrophic** (symbiosis with endomycorrhizal fungi)

Pyrrolizidinic alkaloids content in whole plant parts (nematostatic effect)



- ⇒ **Growth, Biomass & BNF** (^{15}N) of *C. juncea* in vegetable producing tunnels in Mediterranean conditions (France & Spain)
- ⇒ Incidence of *C. juncea* on following crop: **yield, nitrogen supply, nematode protection**
- ⇒ **Effect on soil biological activity**: microbial activity, mycorrhizogenous potential, microbial populations
- ⇒ Symbiosis : **nodulation & survival** of rhizobia in Mediterranean conditions
- ⇒ **Soil** biology, fertility & sanitary improved after 3 years ?

Assay on nematostatic and green manuring effects of *Crotalaria juncea* In tunnel greenhouse vegetable production (lettuce)



Current rotation cropping

Jan-Mar	lettuce
Mar-Jul	Melon
Jul-Sep	Sorghum (intercrop)
Oct-Dec	Lettuce



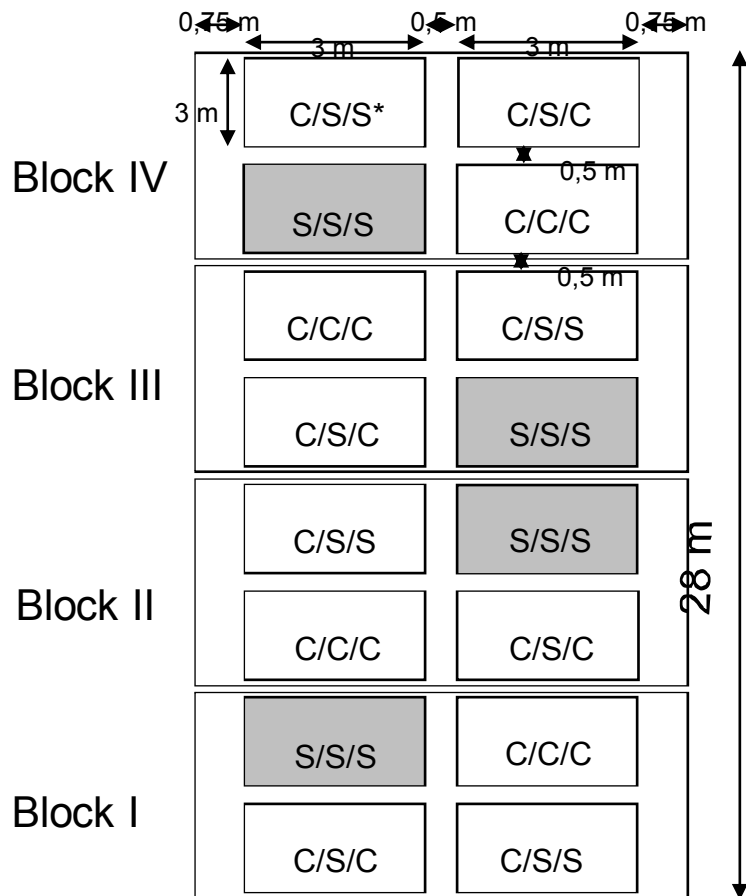
Mr Ginoux (farmer in Noves, Provence)
Mr Deleuze (Delbon)
Dr Prin (CIRAD)



Severe nematode problem

Experimental design over 3 years

4 blocks × 4 treatments



Crotalaria/Sorgho/Crotalaria
 Crotalaria/Crotalaria/Crotalaria
 Crotalaria/Sorgho/Sorgho
 Sorgho/Sorgho/Sorgho

Inoculation
5 strains



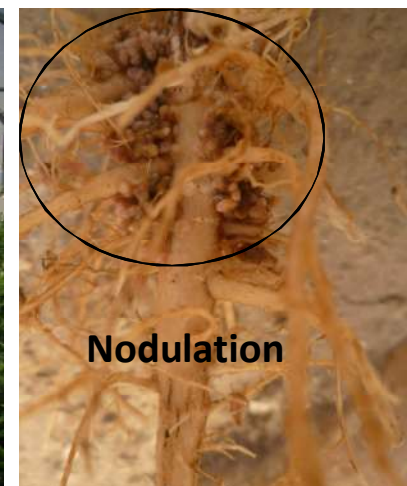
3 experimental sites:
 Noves, Aimargues (France)
 Archena (Spain)

Crotalaria juncea

Crotalaria seedling

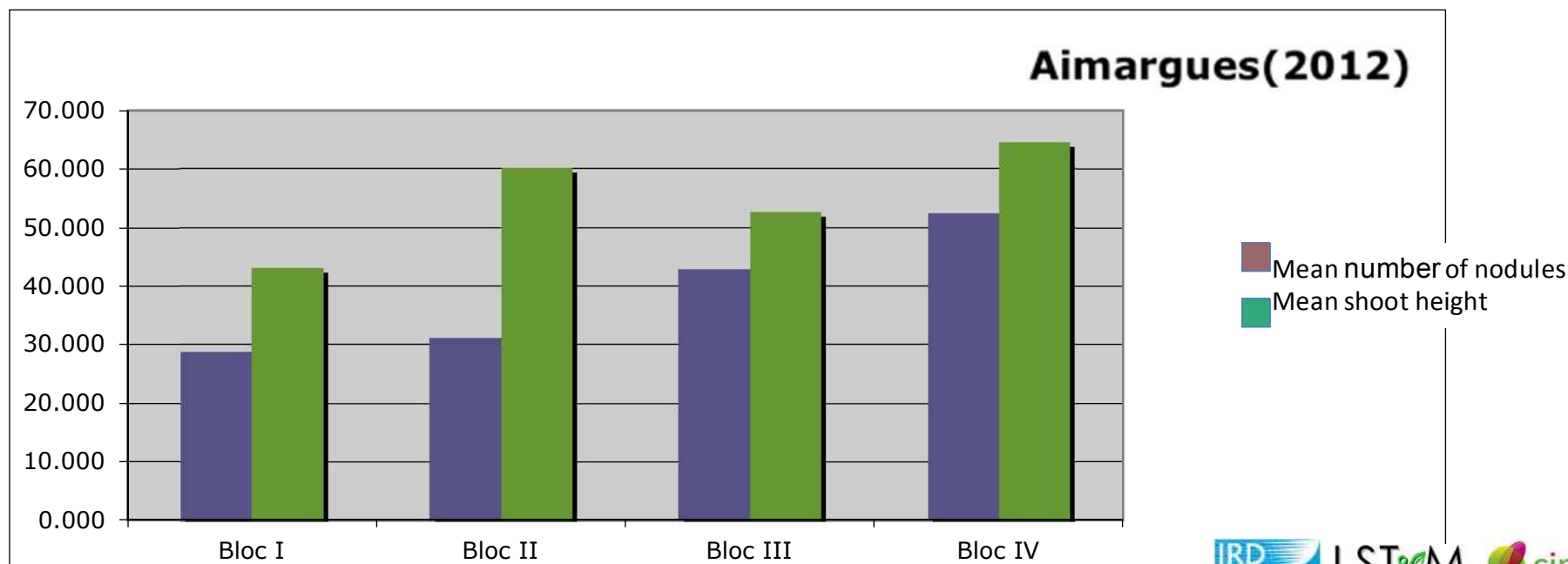
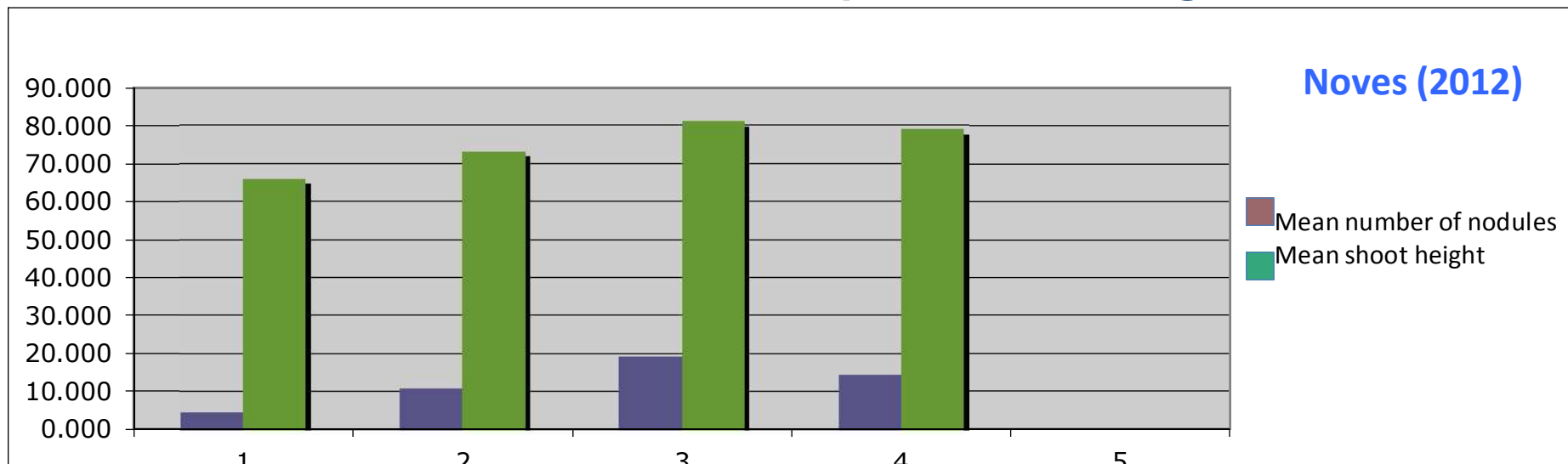


Inoculation



Crotalaria juncea in South France Growth, biomass, nodulation

Plant growth and nodulation of *C. juncea* after 6 weeks growing in the 4 blocks of the experimental designs



Cut, shear, weigh, bury



Lettuce crop



Observation of nematode galls on lettuce roots at harvest (75 days) After *Sorghum* or *Crotalaria*

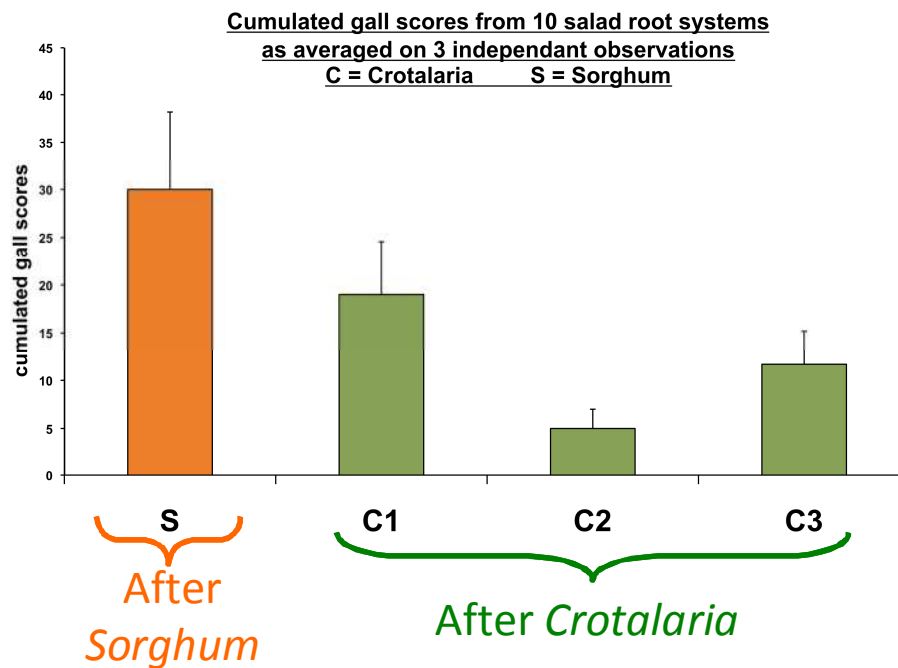


Evaluation of nematode infection on lettuce roots at harvest

Gall index

ANNEXE IV
Indexation du niveau d'attaque par observation des galles

0	Système racinaire complet et sain. Pas d'infection.	6	50 % du système racinaire comporte des galles et ne fonctionne plus.
1	Très peu de petites galles peuvent être détectées après examen.	7	75 % du système racinaire comporte des galles et entraîne une perte de production.
2	De petites galles comme dans le 1 et 2 mais plus nombreuses et facilement détectables.	8	Les racines sont malades, la plante n'a plus de nourriture.
3	Nombreuses petites galles, la croissance et les fonctions de la racine ne sont pas sérieusement affectées.	9	Le système racinaire est plein de galles, la plante est en train de mourir.
4	Nombreuses petites galles, quelques grosses galles. La majorité du système racinaire fonctionne encore.	10	Plantes et racines sont mortes.
5	25 % du système racinaire comporte des galles et ne fonctionne plus.		



→ Significant differences between plots and treatments

Lettuce harvest (fresh weight) Year 1

France Site 1 Year 1

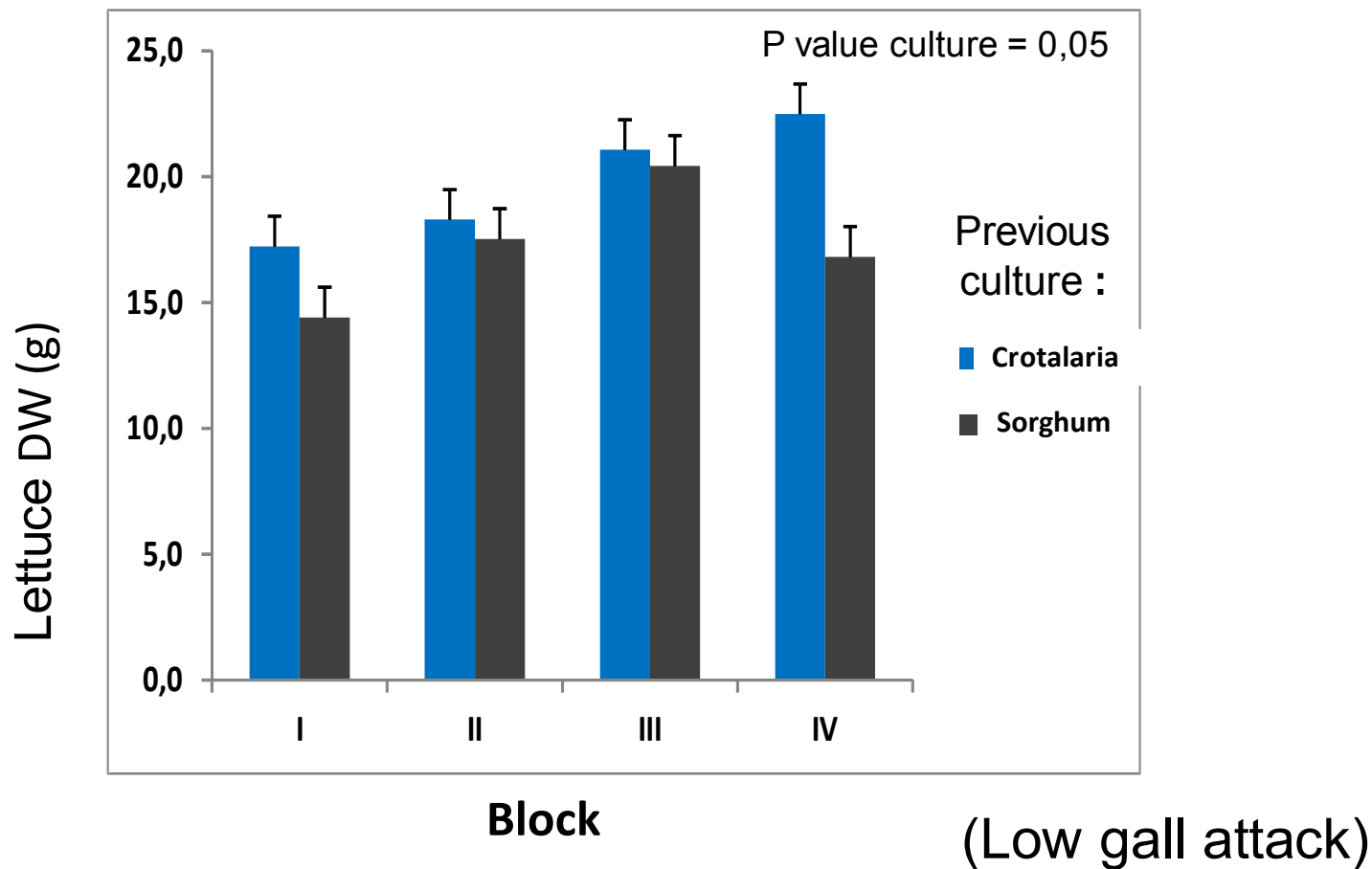
Newman-Keuls (SNK)		P < 0.001
Treatment	Average (g)	Groups
Crotalaria	364	A
Crotalaria	352	A
Crotalaria	352	A
Sorghum	300	B

France Site 2 Year 1

Newman-Keuls (SNK)		P < 0.05
Treatment	Average (g)	Groups
Crotalaria	425	A
Crotalaria	409	AB
Crotalaria	403	AB
Sorghum	376	B

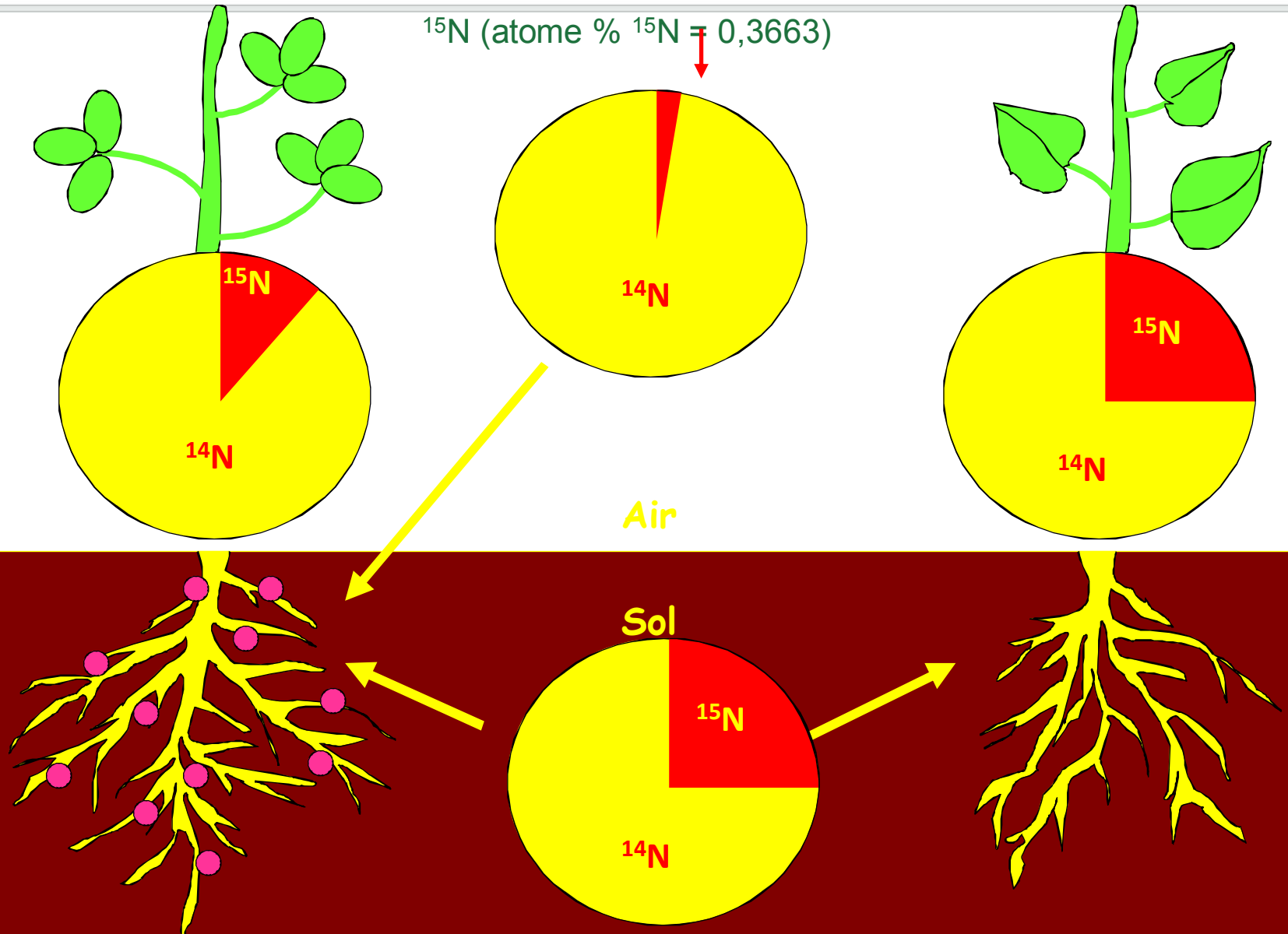
- a block effect
- previous crotalaria improves salad biomass compared to control sorghum

Effect of previous culture *Crotalaria* vs. Sorghum on lettuce biomass (DW) Noves year 2

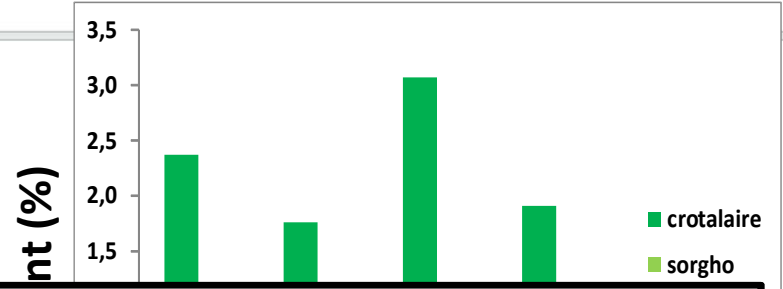
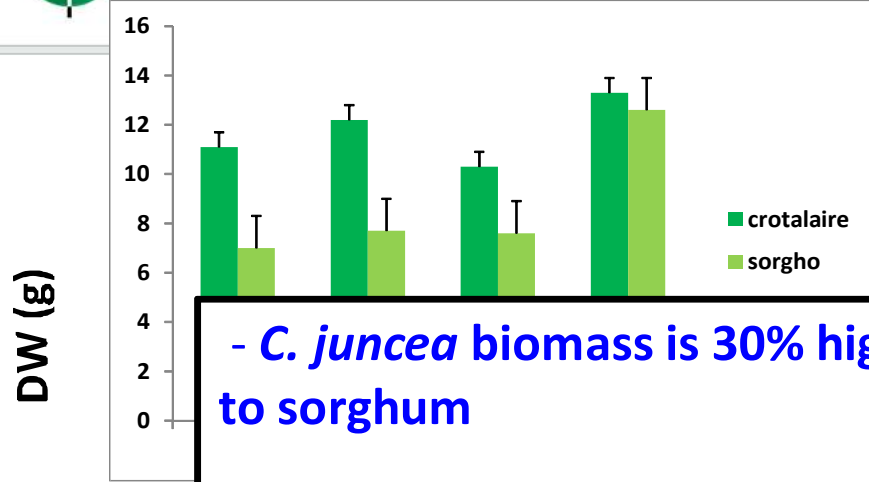


Nitrogen fixation Isotopic approach

Estimation of nitrogen fixation by ^{15}N natural abundance measurement



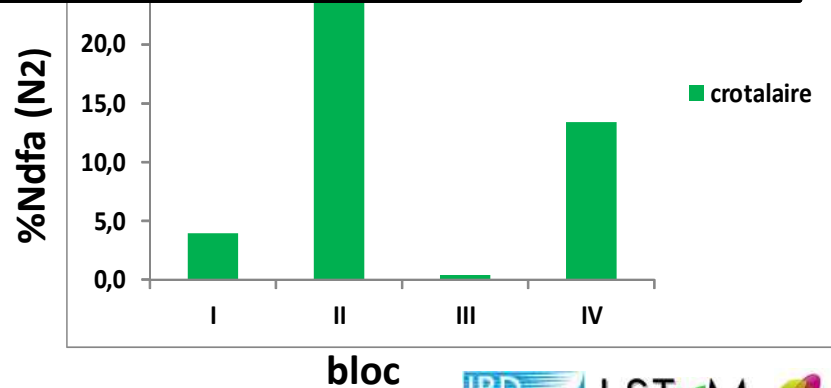
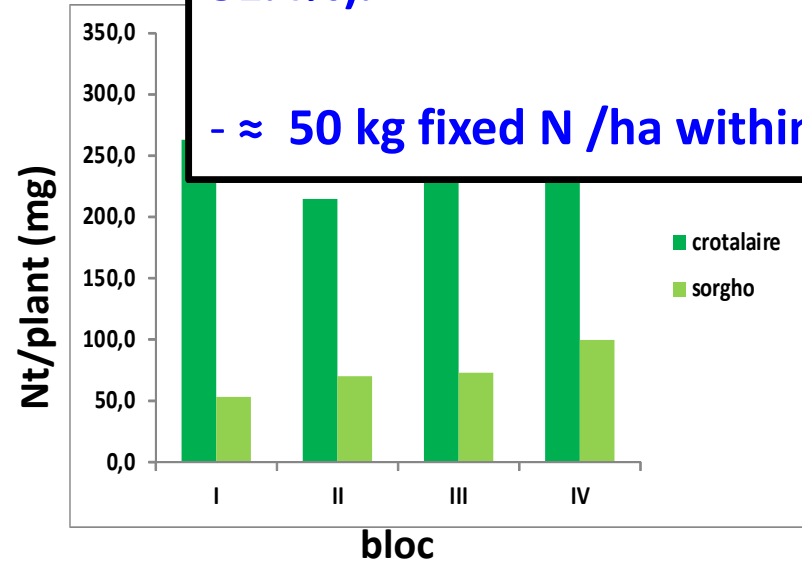
¹⁵N natural abundance, N content & %N₂ fixed in *C. juncea* after 2 month-growth (experimental designs / 4 blocks)



- *C. juncea* biomass is 30% higher with 3-fold N content compared to sorghum

-The part of N₂ fixation ($\Delta^{15}\text{N}$) in *C. juncea* varies in blocks (0.4-31.4%).

- \approx 50 kg fixed N /ha within 3 months. Comparable to bean.



Microbiological approach

Soil microbial activity

Genetic structure of soil bacterial and fungal communities

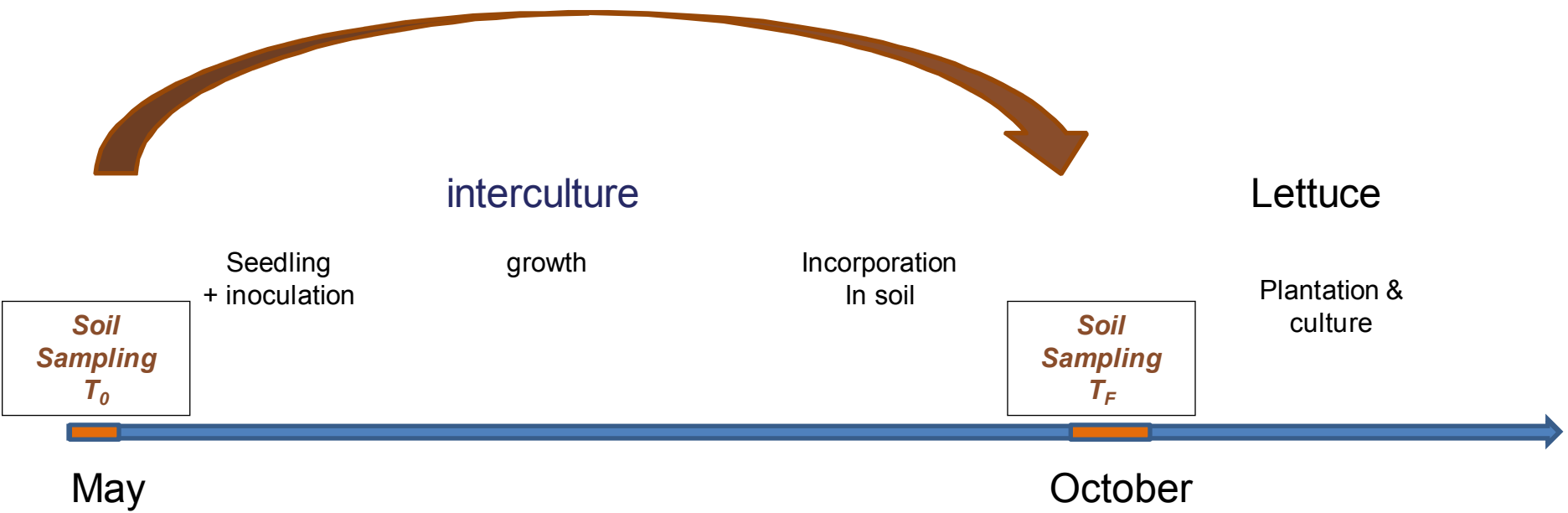
Soil mycorrhizogenous potential

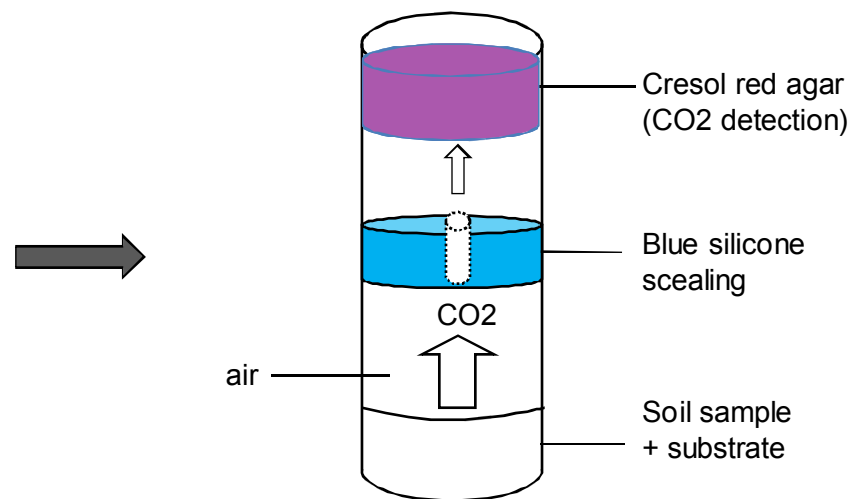
Inoculum survival in soil (MPN)

Inoculum strain efficiency

Noves (France) 2011, 2012

Interculture *Crotalaria juncea* versus *Sorghum bicolor*





sugars

Organic acids

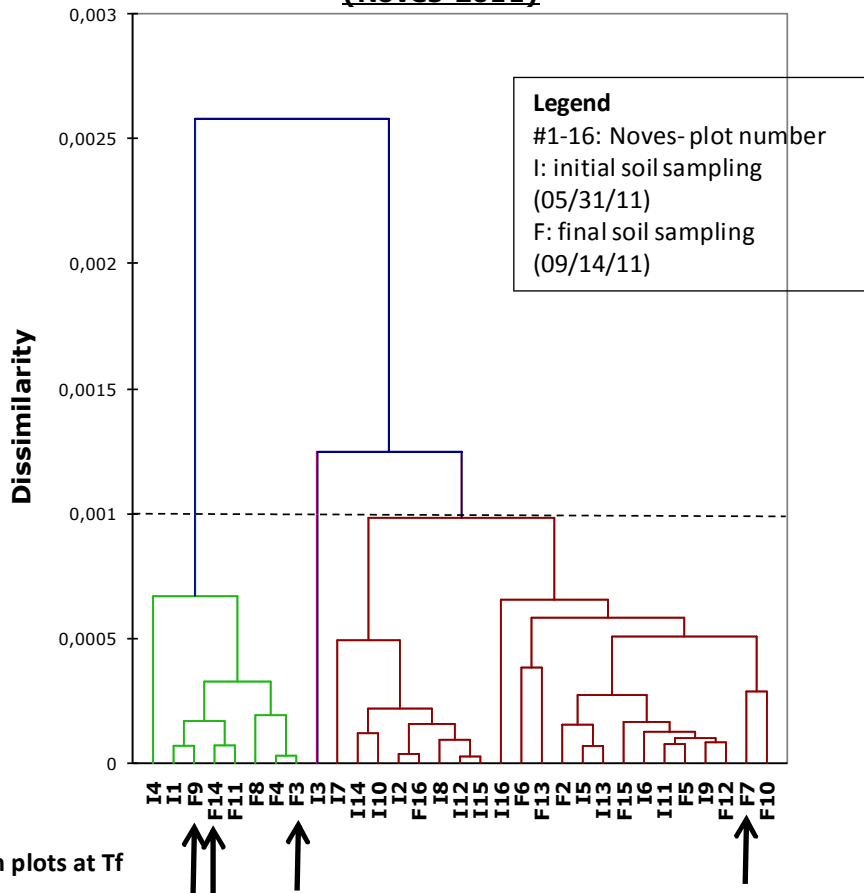
Amino acids

	1	2	3	4	5	6	7	8	9	10	11	12
A	mannose	mannose	mannose	mannitol	mannitol	mannitol	trehalose	trehalose	trehalose	arabinose	arabinose	arabinose
B	H ₂ O	H ₂ O	H ₂ O	xylose	xylose	xylose	sucrose	sucrose	sucrose	galactose	galactose	galactose
C	inositol	inositol	inositol	sorbitol	sorbitol	sorbitol	rhamnose	rhamnose	rhamnose	arabitol	arabitol	arabitol
D	glutamate	glutamate	glutamate	citrate	citrate	citrate	maleate	maleate	maleate	ac.mallique	ac.mallique	ac.mallique
E	gluconate	gluconate	gluconate	ascorbate	ascorbate	ascorbate	etoglutarate	etoglutarate	etoglutarate	D-oxalate	D-oxalate	D-oxalate
F	asparagine	asparagine	asparagine	valine	valine	valine	methionine	methionine	methionine	glutamine	glutamine	glutamine
G	N-acétylglu	N-acétylglu	N-acétylglu	alanine	alanine	alanine	serine	serine	serine	histidine	histidine	histidine
H	proline	proline	proline	leucine	leucine	leucine	lysine	lysine	lysine	arginine	arginine	arginine

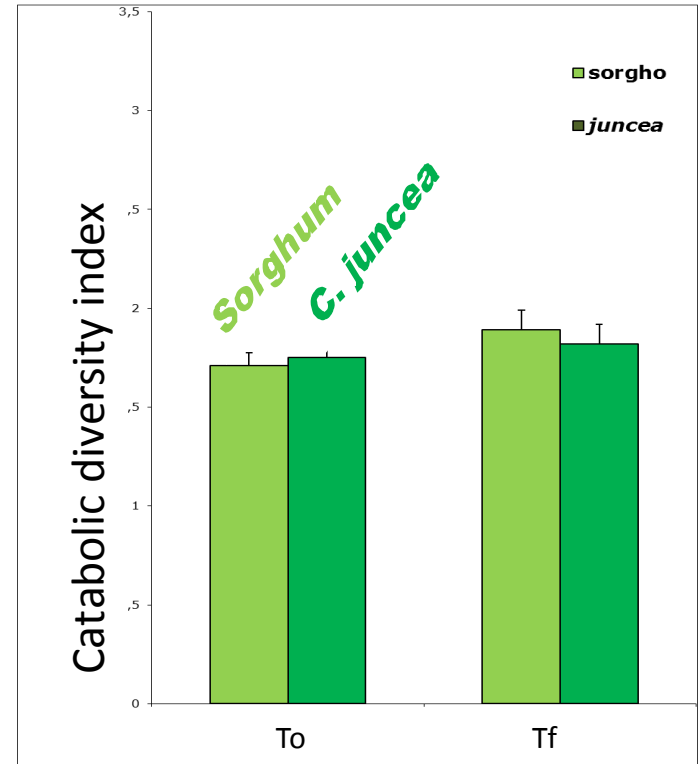
32 substrates
(triplicates)

Soil catabolic profiles To vs. Tf

(Noves-2011)



Cluster analysis of the catabolic profiles characterizing soils sampled before the *C. juncea* seeding and after its soil incorporation and mineralisation.



$$H = - \sum P_i \ln P_i$$

P_i : relative oxydation intensity of a substrate i reference to the total oxydation level of all substrates

Microbiological approach

Soil microbial activity

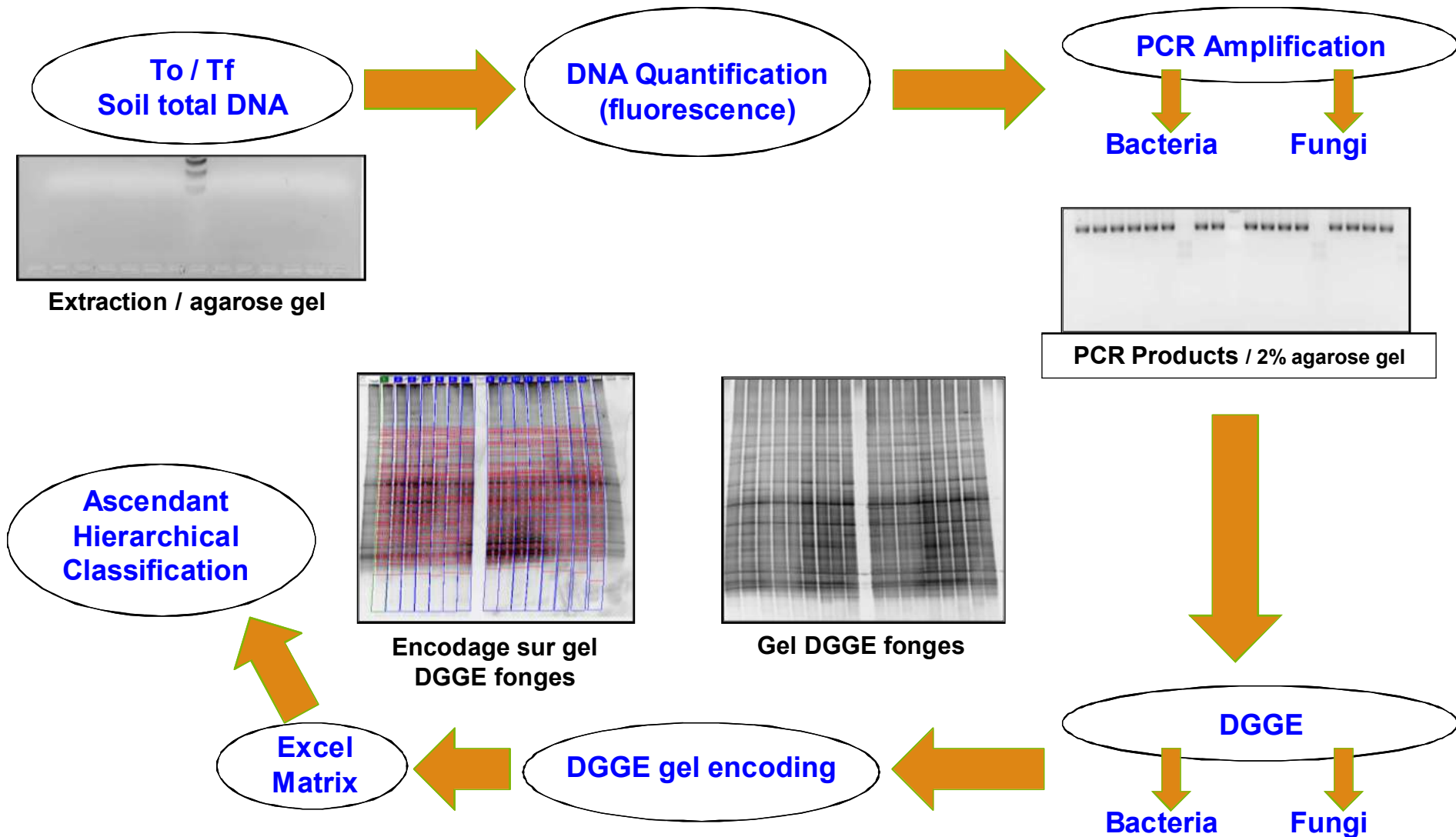
Genetic structure of soil bacterial and fungal communities

Soil mycorrhizogenous potential

Inoculum survival in soil (MPN)

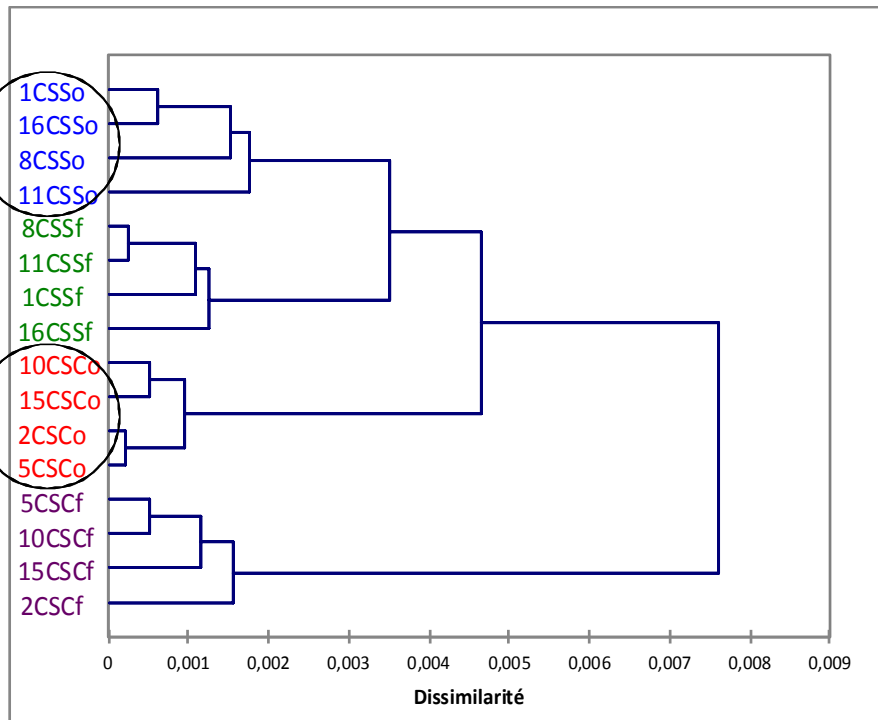
Inoculum strain efficiency

- Total bacteria : 16S rRNA coding gene (200 bp)
- Total fungi : 18S rRNA coding gene (400 bp)

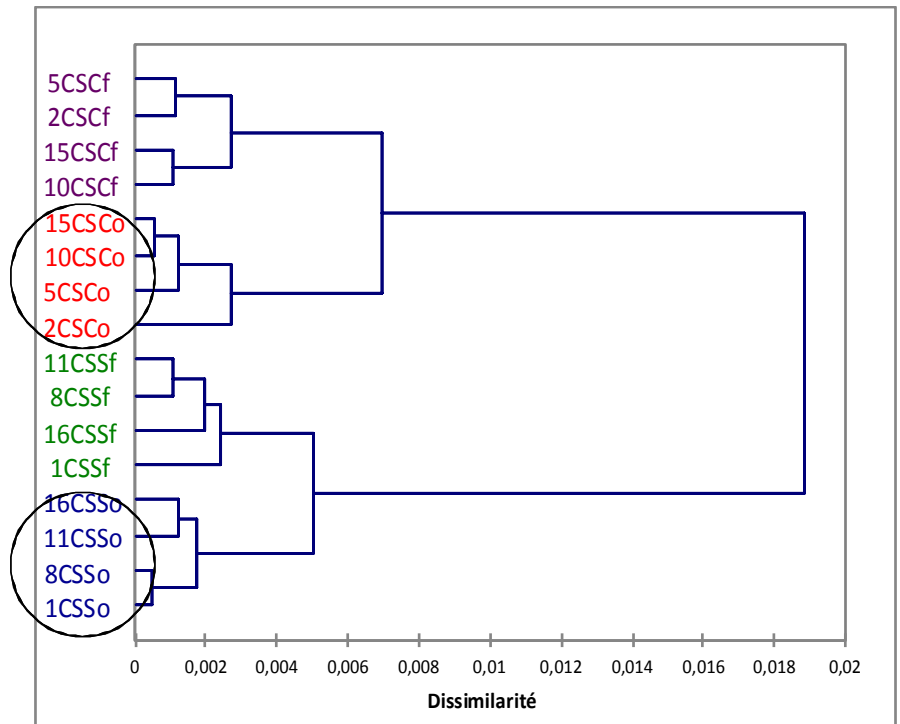


Noves 2011 (1st year)
Crotalaria Plots To

AHC **Bacterial community**

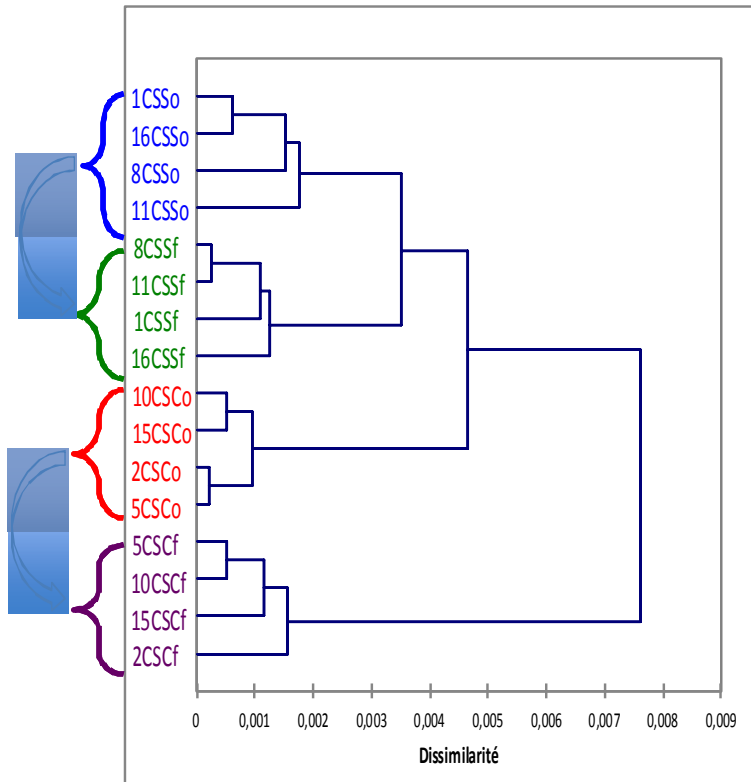


AHC fungal community

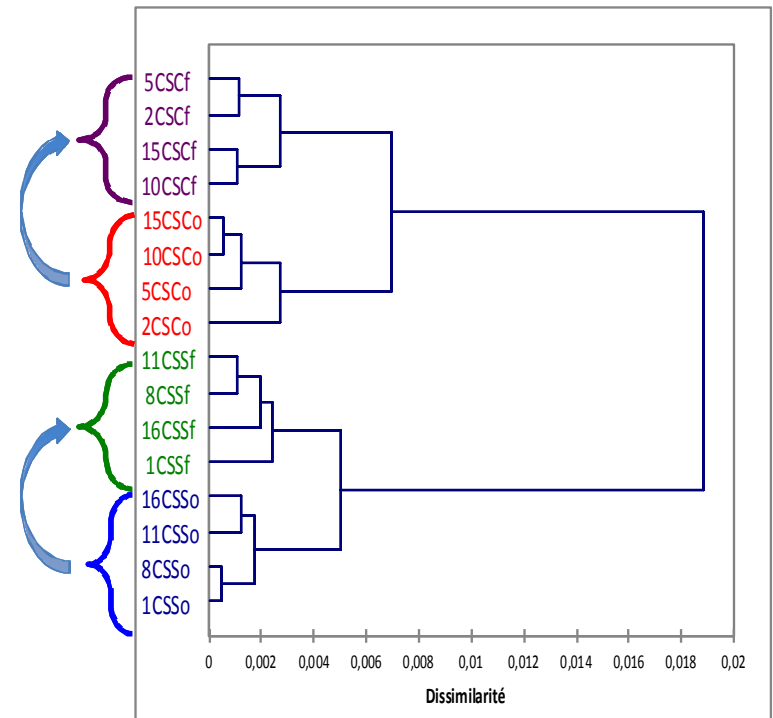


T_0 : initial heterogeneity of plots

AHC Bacterial community



AHC fungal community



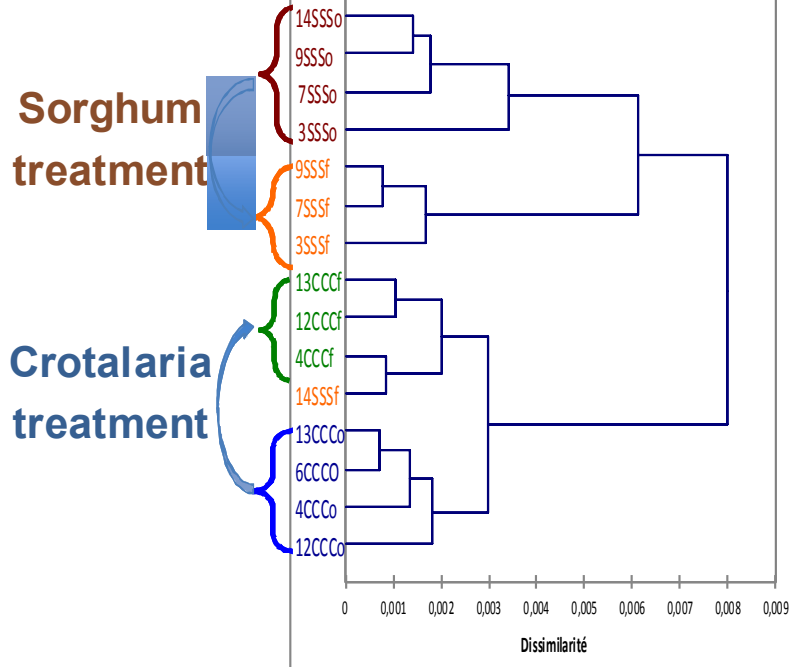
Crotalaria Effect
Tf ≠ To

2011 (1st year)

Crotalaria Plots versus *Sorghum* plots

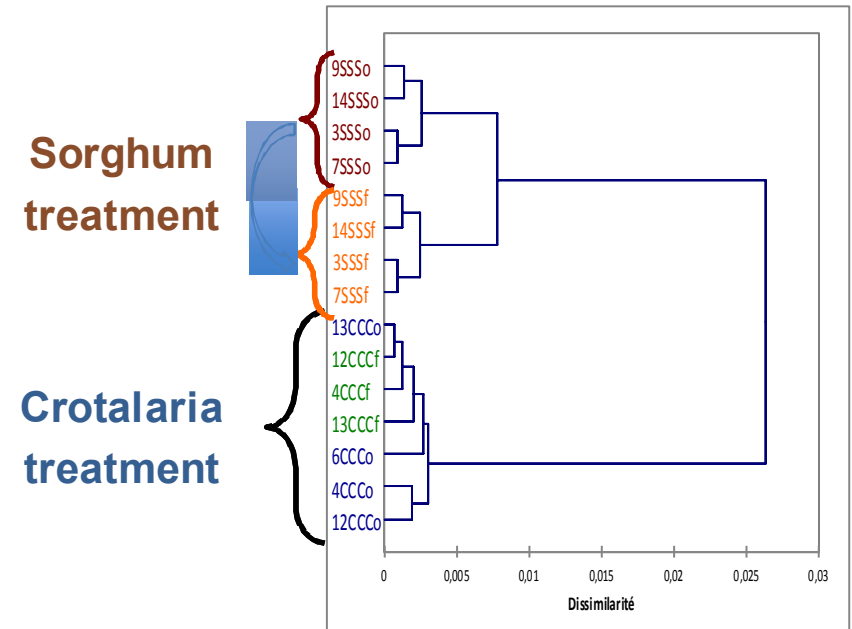
BACTERIA

AHC **Bacterial community**



FUNGI

AHC fungal community



Plant effect
Sorghum* > *Crotalaria

Partial conclusions

Interculture impacts genetic structures of both soil bacterial and fungal communities,

Sorghum ≠ *Crotalaria*

Intensity varying in campainings.

Initial soil heterogeneity persists after intercultures.

Microbiological approach

Soil microbial activity

Soil mycorrhizogenous potential

Inoculum survival in soil (MPN)

Inoculum strain efficiency



Soil mycorrhizogenous infective potential evaluation (t=0) trap culture

Trap cultures of mycorrhizal fungi using *C. juncea* and *L. multiflorum* on soils sampled in the 4 sorghum plots (Noves-tunnel, 2011).




C. juncea



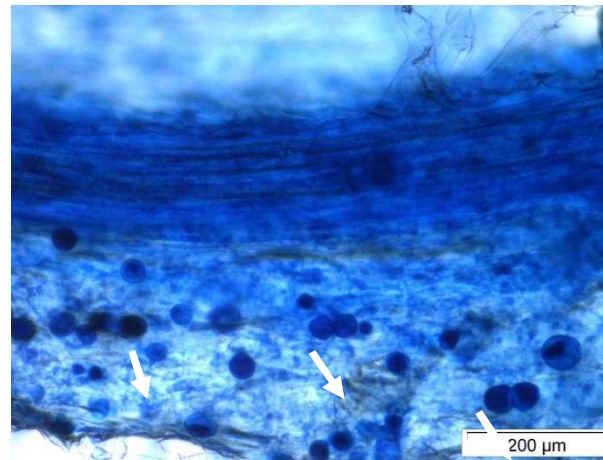
Ray grass

Root systems at harvest.

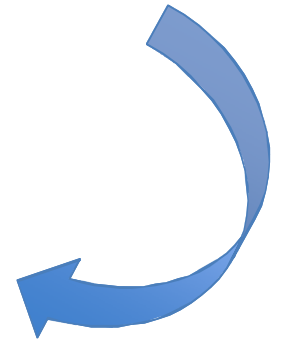
 **Soil mycorrhizogenous infective potential
evaluation (t=0) trap culture**



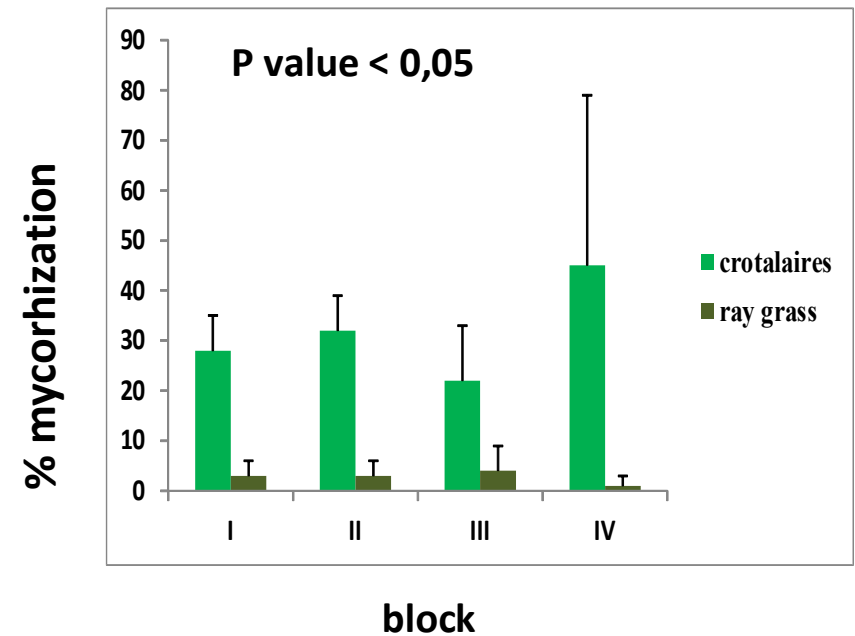
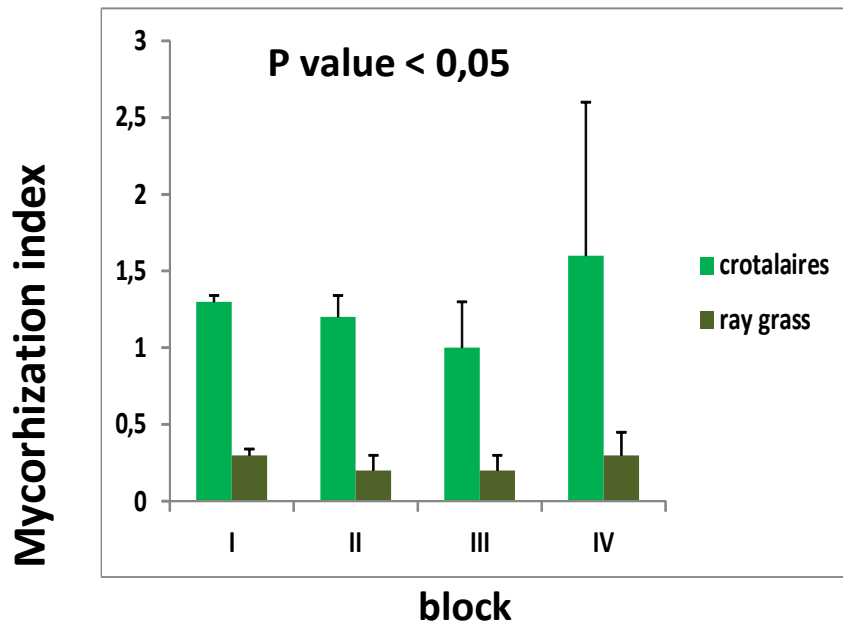
Noves soil +
{45% attapulgate, 45% sand, 10% loam}
subdued 2 mm, 36 day cultivation



Microscopic observation of inner root
mycorrhizal structures from Trypan Blue stained roots



Soil mycorrhizogenous infective potential evaluation (t=0) trap culture



Crotalaria >> Ray grass
C. juncea may monitor soil mycorrhizal infectivity

Microbiological approach

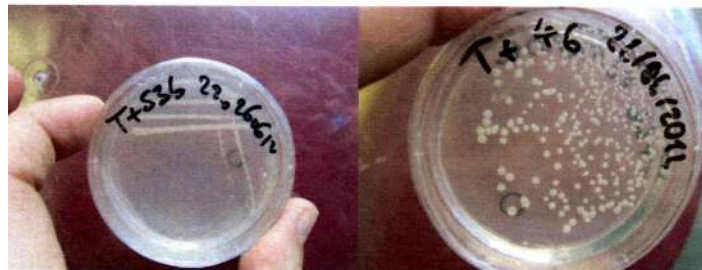
Soil microbial activity

Soil mycorrhizogenous potential

Inoculum survival in soil (MPN)

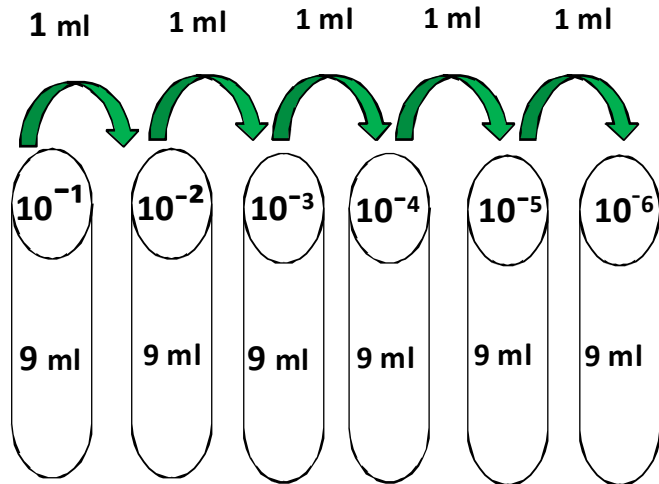
Inoculum strain efficiency

Rhizobial trapping after 1 year





Rhizobial inoculum persistence after *C. juncea* cultivation (MPN)



× 4 C/C/C soil samples × 4 repeats

$$X = (m*d)/v$$



X – estimated number rhizobiums/g soil

m –bacterial number at the lowest dilution

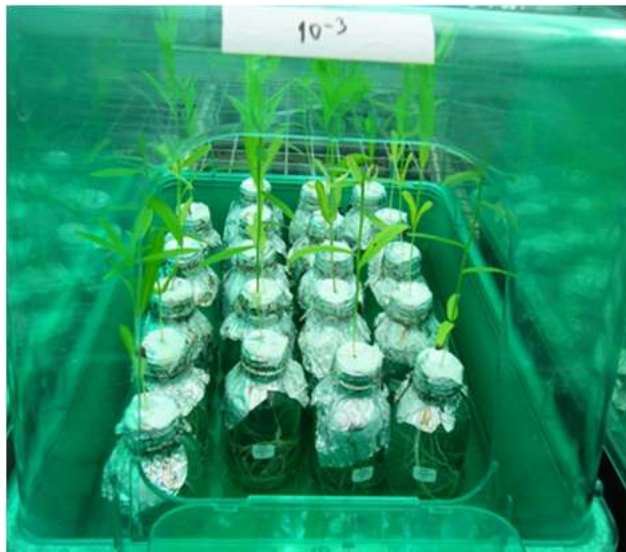
d – the lowest dilution

v –inoculum volume (= 1 ml)

bloc	I					II					III					IV				
	1	2	3	4	total	1	2	3	4	total	1	2	3	4	total	1	2	3	4	total
répétition																				
dilution																				
10 ⁻¹	+	+	+	+	4	+	+	+	+	4	+	+	+	+	4	+	+	+	+	4
10 ⁻²	+	+	-	+	3	-	+	+	-	2	+	+	+	+	4	+	+	+	-	3
10 ⁻³	-	+	-	-	1	+	-	-	-	1	+	+	+	+	4	-	-	-	+	1
10 ⁻⁴	+	+	-	-	3	-	-	+	+	2	+	+	+	+	4	+	-	+	+	3
10 ⁻⁵	-	-	+	+	1	-	+	-	-	1	-	+	-	-	1	-	-	+	-	1
10 ⁻⁶	-	-	-	-	0	+	+	-	-	2	+	+	+	-	3	-	-	-	-	0
total/bloc					12					13					20					12



x = 1,7 * 10³ bact /g soil



Microbiological approach

Soil microbial activity

Soil mycorrhizogenous potential

Inoculum survival in soil (MPN)

Inoculum strain efficiency



West Africa Crotalaria nitrogen fixing bacterial symbionts

Long Bacilli

Slow growth

Bradyrhizobium spp.

(Samba *et al.*, 1999)

Short Bacilli

Fast growth

Methylobacterium nodulans

Methanol +

Formate +

Formaldehyde +

Methylamine -

nodA, nifH +

(Sy *et al.*, 2001

Jourand *et al.*, 2004)



Sterile substrate :
45% perlite, 45% sand,
10 % loam
+ N-free nutritive solution
Analysis 30 dai.



Bradyrhizobium

ORS 1816 (origin *C. hyssopifolia*)

ORS 1815 (origin *C. hyssopifolia*)

ORS 1810 (origin *C. lathyroides*)

ORS 1929 (origin *C. comosa*)

ORS 1935 (origin *C. gorensis*)

Methylobacterium nodulans

ORS 2060 (origin *C. podocarpa*)

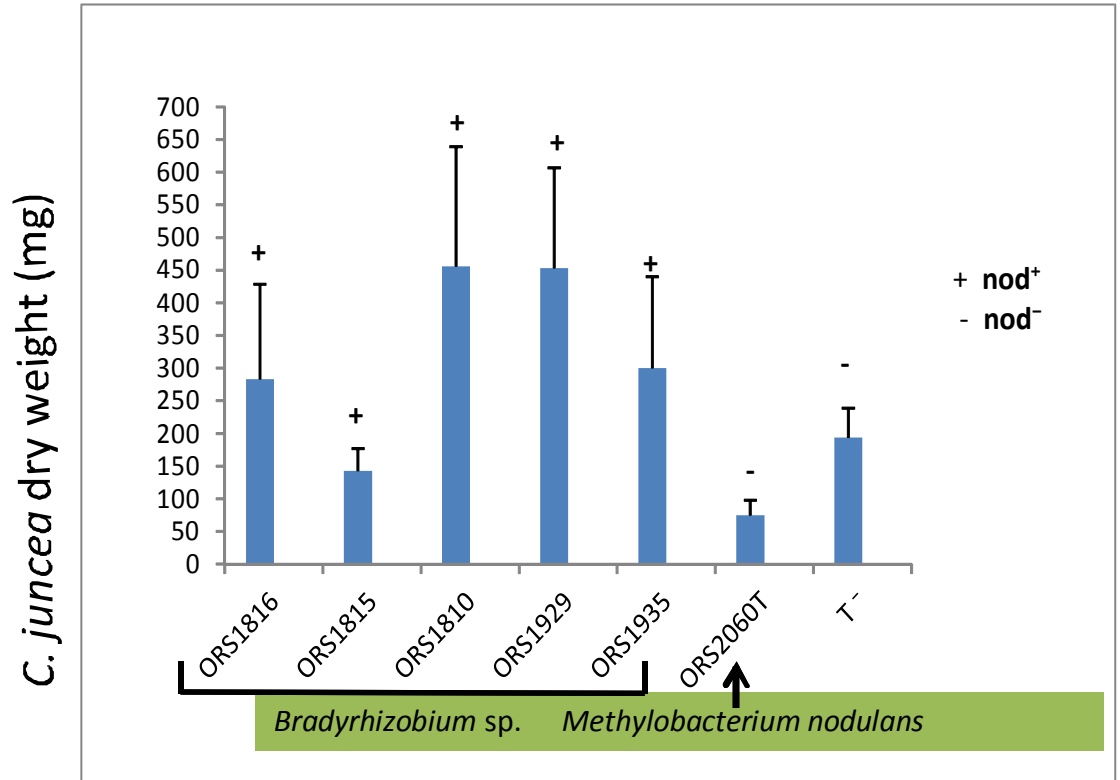


Efficiency potential of the 5 *Bradyrhizobium* Inoculant strains on *C. juncea*



Contrasting strains

P value < 0,05



Molecular approach

Inoculant Rhizobial strain molecular characterization & tracing in soil and time

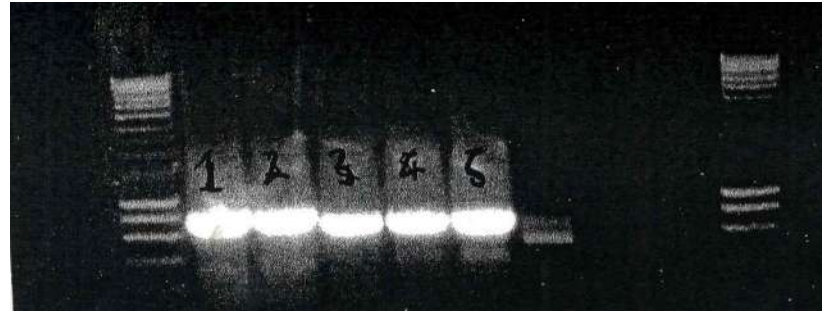


5 inoculant strains

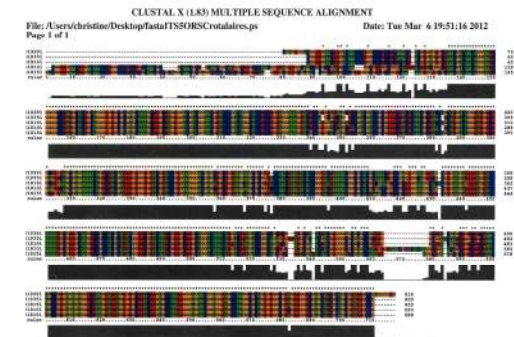
16S-23S ITS
PCR-Amplification

ORS 1816
ORS 1929
ORS 1810
ORS 1935
ORS 1815

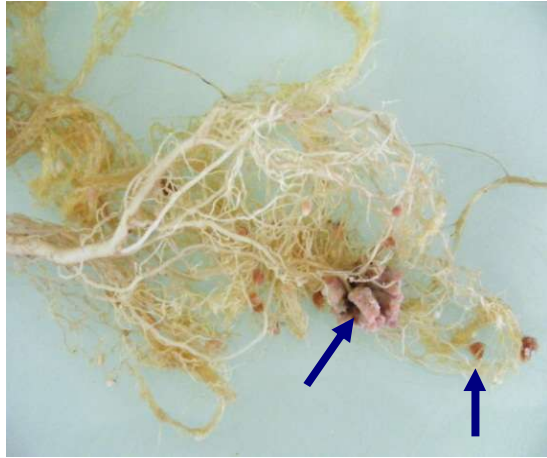
800pb
600pb



- The 5 strains are *Bradyrhizobium* spp.
- ITS polymorphism among the inoculant strains
- No natural *Crotalaria* nodulating strain in experimental soils



Noves, 1 year later...



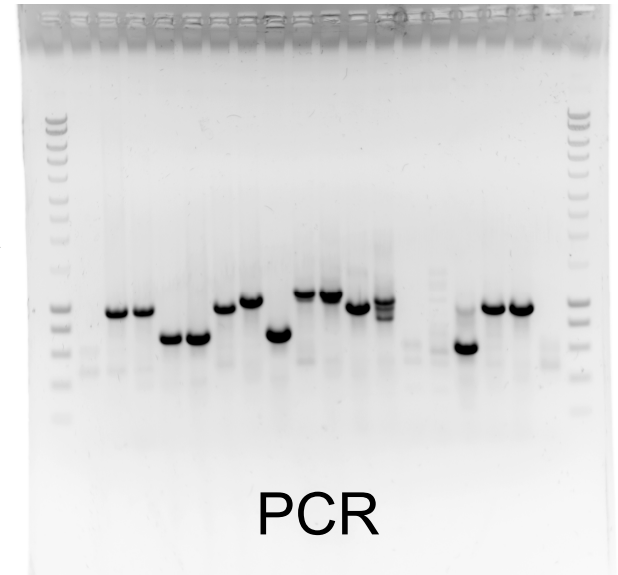
C. juncea nodules
(Noves samples)



Bacterial
isolation



DNA extraction

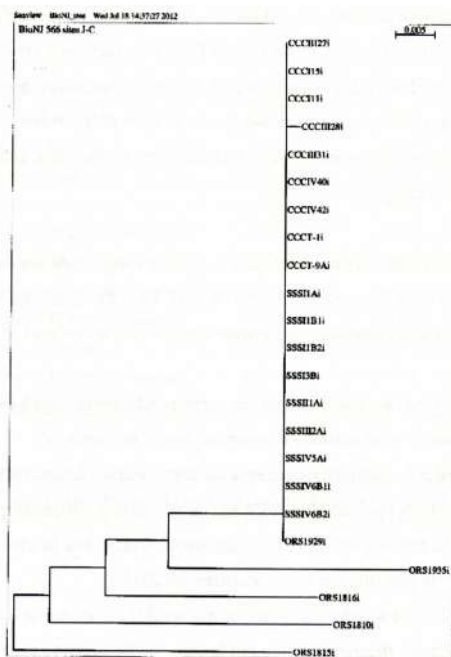
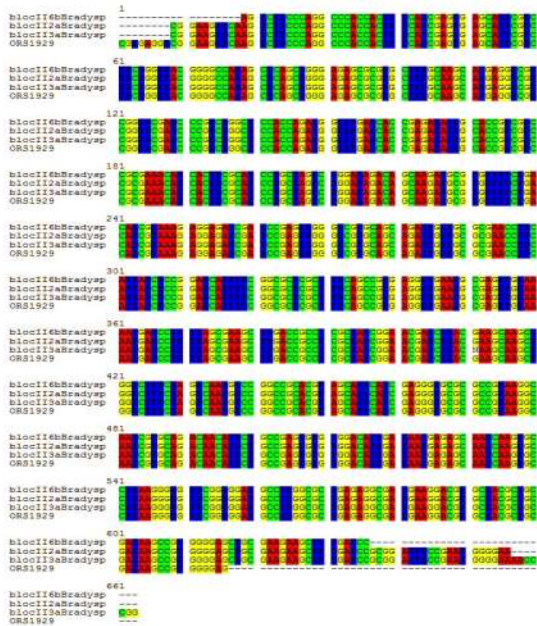


PCR

Bradyrhizobium sp
A. radiobacter
Rhizobium spp

16S-23S ITS rDNA
sequencing

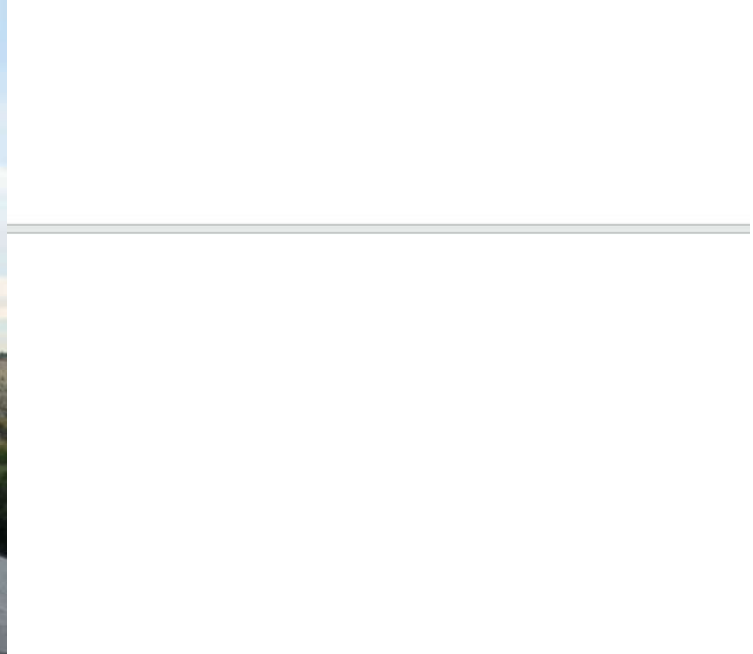




Noves & Archena

After 1 year & in all plots :
 100% nodule *Bradyrhizobium*
 isolates identical to ORS 1929

ORS 1929 = sole strain able
 to survive ?
 Most competitive ?



Archena (Spain)
Assay in a Canarian shelter



C. juncea

Archena (Spain) Assay in a Canarian shelter



Nodules



Archena
tomato after *Crotalaria*
(green manure)





↑
Archena
Tomato &
Crotalaria
associated
cultures



Galls on tomato roots

Harvest daily registration

2012 (Year 1) & 2013 (Year 2)

- *Crotalaria* / tomato rotation
- *Crotalaria* + tomato co-culture



Half yield compared to traditional
Good quality tomato fruits

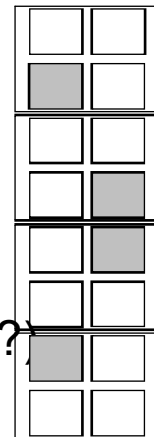
Farmer

Take home message

- ⇒ *C. juncea* in Mediterranean tunnel conditions exhibits a **rapid growth** with **abundant biomass** (> Sorghum)
- ⇒ The biological nitrogen fixing potential of *C. juncea* may be similar to bean (**50 Kg N /ha / 3 months**)
- ⇒ Positive **Green Manure effect** of *Crotalaria juncea* on lettuce biomass (+ **10%**)
- ⇒ **Reduced gall attack** observed after *Crotalaria* (lettuce/tomatoe ; France & Spain)
- ⇒ **Soil mycorrhizogenous potential** may be higher after *C. juncea* (>>> ray grass)
- ⇒ *Bradyrhizobium* spp. inoculant strains may **persist and fix N₂** in mediterranean conditions (France & Spain)
 - ⇒ Impact of *C. juncea* culture on **soil bacterial / fungal community**
 - ⇒ Nitrogen transfer to following crop ? (in progress over 3 years)
 - ⇒ Soil chemical composition / fertility after 3 years ?

After 3 years in Noves (France)

- Vegetable production is more **homogenous**, more **abundant**, **good quality**.
- In year 2 and 3 (2012 et 2013) all **surrounding greenhouses** faced **caterpillar** & nematode attacks, but not the trial greenhouse: *Crotalaria* might have a **zonal effect** larger than the unit plot.
- **Convinced** to go on / switch from *Sorghum* to *Crotalaria* interculture (seeds ?)



After 2 years in Archena (Spain)

- **Surprised** to harvest good quality tomatoes, even half yield
- (Year -1 = 0 harvest)
- *Crotalaria* = **barrier** effect against nematodes ?
- Year 3 : in progress



Christine Le Roux
Marc Ducouso
Yves Prin Y.,
Antoine Galiana



Philippe Jourand
Julie Bourrillon
Ezekiel Baudoin
Pierre Tisseyre
Marc Boursot
Robin Duponnois
Philippe de Lajudie



François Deleuze
José Picon
<http://www.delbon.com>



Florian Carlet
Eric De Belder
Julien Caillère



Natalia Kulagina
Thibaud Darré,
Damien Cécillon



Legume-supported cropping systems for Europe (Legume Futures)
is a collaborative research project funded from the European Union's Seventh Programme
for research, technological development and demonstration under grant number 245216